

Using DNA metabarcoding for assessing chironomid diversity and community change in mosquito controlled temporary wetlands

Kathrin Theissinger¹, Anna Kästel¹, Vasco Elbrecht², Jenny Makkonen^{1,3}, Susanne Michiels⁴, Susanne I. Schmidt¹, Stefanie Allgeier¹, Florian Leese², Carsten A. Brühl¹

¹ University of Koblenz-Landau, Institute for Environmental Sciences, Landau, Germany.

² University of Duisburg-Essen, Aquatic Ecosystem Research, Essen, Germany.

³ University of Eastern Finland, Department of Environmental and Biological Sciences, Kuopio, Finland.

⁴ AquaDiptera, Emmendingen, Germany.

Corresponding author: Kathrin Theissinger (theissinger@uni-landau.de)

Academic editor: Kristy Deiner | Received 18 September 2017 | Accepted 21 December 2017 | Published 15 January 2018

Abstract

The biocide *Bacillus thuringiensis* var. *israelensis* (Bti) is widely applied for mosquito control in temporary wetlands of the German Upper Rhine Valley. Even though Bti is considered environmentally friendly, several studies have shown non-target effects on chironomids, a key food resource in wetland ecosystems. Chironomids have been proposed as important indicators for monitoring freshwater ecosystems, however, morphological determination is very challenging. In this study, we investigated the effectiveness of metabarcoding for chironomid diversity assessment and tested the retrieved chironomid operational taxonomic units (OTUs) for possible changes in relative abundance and species diversity in relation to mosquito control actions in four temporary wetlands. Three of these wetlands were, for the first year after 20 years of Bti treatment, partly left Bti-untreated in a split field design, and one wetland has never been treated with Bti. Our metabarcoding approach detected 54 chironomid OTUs across all study sites, of which almost 70% could be identified to species level comparisons against the BOLD database. We showed that metabarcoding increased chironomid species determination by 70%. However, we found only minor significant effects of Bti on the chironomid community composition, even though Bti reduced chironomid emergence by 65%. This could be due to a time lag of chironomid recolonization, since the study year was the first year of Bti intermittence after about 20 years of Bti application in the study area. Subsequent studies will have to address if and how the chironomid community composition will recover further in the now Bti-untreated temporary wetlands to assess effects of Bti.

Key Words

Bacillus thuringensis var. *israelensis* (Bti), Barcode of Life Database (BOLD), chironomid saprobic index, cytochrome oxidase subunit I (COI), ecotoxicology, macrozoobenthos emergence, operational taxonomic units (OTU)

Introduction

Since 1981, the biocide *Bacillus thuringiensis* var. *israelensis* (Bti) is widely applied for mosquito (Culicidae, Diptera) control in temporary wetlands of the German Upper Rhine valley to minimize nuisance of local residents (Becker 1998). Bti is considered as the most environmentally friendly alternative to chemical pesticides for mosquito control due to a supposedly high specificity to mosquito larvae and negligible non-target effects even on closely related dipterans (Boisvert and Boisvert 2000).

This is important as large areas of both aquatic and terrestrial habitats of the Upper Rhine valley are protected (bird sanctuaries, nature reserves and Natura 2000 sites) and comprise of biodiversity hotspots (Biggs et al. 2005, Lukács et al. 2013).

However, several studies have shown that Bti non-target effects are possible (reviewed in Boisvert and Boisvert (2000)). Non-biting midges (Chironomidae, Diptera) are the most Bti-sensitive non-target family (Boisvert and Boisvert 2000). Controlled experiments revealed varying mortality rates on chironomid larvae with older larvae

being typically less sensitive to Bti (Ali et al. 1981, Treverrow 1985, Ping et al. 2005). They also reported different sensitivities among species (Yiallouros et al. 1999) and subfamilies (Liber et al. 1998). A recent study found that first instar larvae of *Chironomus riparius* are highly susceptible to Bti treatment even at commonly used mosquito control application rates (Kästel et al. 2017). Consequently, Bti application might overproportionately affect chironomid species in early larval stages at the time of application. So far, field studies have yielded ambiguous data on possible side effects of Bti on chironomid abundances. These range from positive effects on chironomid larvae richness possibly due to reduced mosquito competition (Lundström et al. 2010), over no effect on chironomid abundance (Lagadic et al. 2016), to a 35–80% reduction of chironomids abundances (Rodcharoen et al. 1991, Hershey et al. 1995, Vaughan et al. 2008, Poulin et al. 2010, Jakob and Poulin 2016).

Chironomids are a taxonomically and ecologically highly diverse group and often dominate all kinds of lotic and lentic ecosystems in terms of species abundances and biomass (Ferrington 2008). With sometimes over 50% of the total macroinvertebrate fauna in aquatic ecosystems (Milošević et al. 2013, Puntí et al. 2009) chironomids are thus a key food resource in wetland ecosystems. They also constitute a central link between aquatic and terrestrial food webs as adult midges are prey for birds, bats, spiders and adult dragonflies (Niemi et al. 1999, Stav et al. 2005, Poulin et al. 2010, Pfitzner et al. 2015). Furthermore, temporal and spatial variability in chironomid community composition has been observed (Lindegård and Brodersen 1995, Rossaro et al. 2006, Milošević et al. 2013) together with a high adaptability of the community to changing environmental conditions (Raunio et al. 2011). Given these particular ecological characteristics, chironomids have been proposed as important indicators for monitoring freshwater ecosystems (Moog 2002, Sæther 1979). However, their morphological determination is very challenging and the taxonomic expertise needed for species identification of chironomids is often lacking (Batzer and Boix 2016). This makes it difficult to study changes in chironomid composition and utilize this as a monitoring tool. DNA-based determination approaches such as DNA barcoding seem therefore promising to support and complement the taxonomic assessment of chironomid community composition.

During recent years, DNA metabarcoding of whole communities has become a new powerful tool for environmental monitoring of aquatic ecosystems (Hajibabaei et al. 2011, Carew et al. 2013, Elbrecht and Leese 2015, Gibson et al. 2015). The DNA-based assays to monitor species biodiversity proved to be a rapid and efficient tool that allows the recovery of a substantial amount of taxa (Sweeney et al. 2011, Taberlet et al. 2012, Yu et al. 2012, Carew et al. 2013, Elbrecht et al. 2017). For metabarcoding in animals, typically the mitochondrial cytochrome oxidase subunit I (COI) is used (Hebert et al. 2003). The COI barcoding region has usually a good tax-

onomic resolution and comparatively well-curated databases as reference for many taxa (Sweeney et al. 2011, Elbrecht et al. 2017). The species present in the sample are identified based on a comparison of retrieved COI sequences (summarized as operational taxonomic units; OTUs) with reference databases (e.g. NCBI or BOLD; Ratnasingham and Hebert 2007). Good species coverage in the database is necessary for taxonomic assignment from sequences, but the identification rate for the different taxa vary widely (Ekrem et al. 2007, Kwong et al. 2012). For Chironomidae only about 30% of the estimated 700 different species in Germany (Samietz 1996) have an entry in the public databases BOLD with formal barcodes (accessed on 3. October 2017, search terms: chironomid & Germany). However, common taxa might be well represented.

Objectives, concept and approach

In this study, DNA metabarcoding was applied to assess the distribution and species richness of chironomids in Bti-treated vs. first year Bti-untreated temporary wetlands in the Upper Rhine Valley. The study sites were part of a mosquito control area that has received regular Bti treatments for approximately 20 years (http://www.kabsev.de/1/1_2/1_2_1/index.php, accessed on 11. August 2017). Our first aim was to study the effectiveness of metabarcoding for chironomid diversity assessment as important and often overlooked freshwater bioindicator. We expected to obtain more species-level identifications based on molecular methods as compared to traditional taxonomic determination and used these data to calculate the saprobic index for the respective sites. Our second aim was to test for possible changes in the chironomid community composition of the temporary wetlands in response to mosquito control actions. Based on the above-mentioned studies we expected:

- an overall reduction in chironomid abundance at Bti-treated sites,
- a reduction in species richness of chironomids at Bti-treated sites, and therefore:
- an overall effect of Bti-treatment on chironomid community composition.

Methodology

Study sites

The study was conducted in Rhineland-Palatinate in southwest Germany close to Neustadt-Geinsheim (Fig. 1). The study sites are regularly flooded in spring and dry out in summer. Thus, the area can be classified as seasonal (= temporary) wetland, which is moreover partly protected as a key amphibian breeding area in the region (Williams 2006). The area has been subject to regular mosquito control management actions for over 20

years, with usually one to two helicopter-applications of Bti between March and June, depending on temperature and precipitation. The study sites Fig. 1 were: “Stiftungsfläche” (S): mainly flooded grassland with some small permanent water bodies, “Großwald” (G): alder carr with larger permanent water bodies, “Mitteltrumm” (M): alder/oak carr with some deeper trenches and ditches and flat sinks. Additionally, the site “Lachen-Speyerdorf” (CL; see Fig. 1) served as control site and was located approximately 7 km away from the sites S, G and M. The site CL was dominated by open alder and pine forest with an abandoned river course.

For the first time after 20 years of regular Bti treatment, parts of the study area were left Bti-untreated in spring 2013 allowing for a split field design. Accordingly, S, G and M were divided into Bti-treated (T; 20 years treated) and untreated (U; first season untreated) site pairs, and CL served as control site never been treated with Bti. The helicopter application took place on April 10, 2013 using IcyPearls (Vectobac WG, ValentBiosciences) at a concentration of 1.44×10^9 ITU/ ha.

Emergence data

Insect imagines were collected weekly with emergence traps ($N = 5$ per site and treatment, in total 35 traps, 0.25 m² area each) over a period of four months (April – July 2013) for 13 weeks after application (WAA; WAA 1 – WAA 13) of Bti. The preserved emergence was deter-

mined to order level and the order Diptera to family level using a Leica M80 microscope and a 10x magnification and counted per trap and sampling week. All chironomid specimens were selected for further analyses. All specimens were conserved in 70% ethanol and stored at room temperature for up to two years until DNA extraction.

Chironomid samples of all emergence traps per WAA were pooled for Bti-treated and Bti-untreated sites. For specific emergence peaks ($N = 18$, see Fig. 2 and text in results) these pooled samples were selected for metabarcoding to determine whether abundance differences can be attributed to a shift in species community composition.

Laboratory methods

Chironomids of emergence peaks ($N = 18$) were selected based on taxonomy and dried overnight at 60 °C. The specimens of each sample were grinded using a Tissue Lyser II (Qiagen, Hilden, Germany) at 30 Hz for 3 × 1 min with two sterile metal beads (3 mm, Hobbyfix, Opitec, Giebelstadt) with a brief centrifugation in between. DNA was extracted following a high salt DNA extraction protocol after (Aljanabi and Martinez 1997). Extraction success was verified using a Nanodrop (ND-1000 Spectrophotometer, Wilmington, USA). 50 µL of DNA from each sample were treated with 1.1 µL RNase (10 mg/mL, Roth, Karlsruhe, Germany) at 37 °C for 30 min, followed by purification using a MinElute Reaction Clean up Kit (Qiagen, Hilden, Germany). The DNA con-

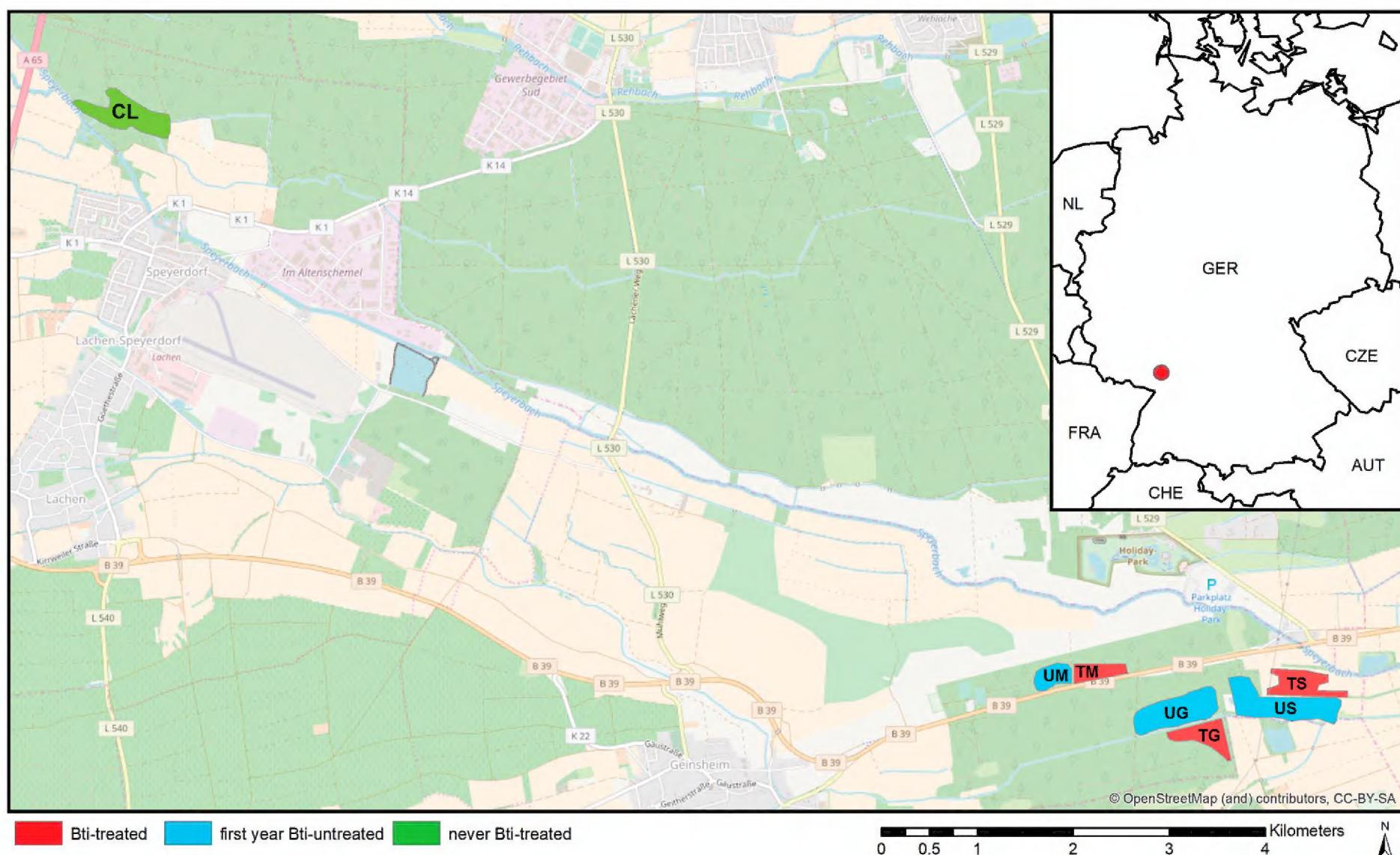


Figure 1. Study sites in southwest Germany close to Neustadt-Geinsheim. “Stiftungsfläche” (S), “Großwald” (G), “Mitteltrumm” (M) “Lachen-Speyerdorf” (CL). S, G and M were divided into Bti-treated (T; 20 years treated) and Bti-untreated (U; first season untreated) site pairs, and CL served as control site never been treated with Bti.

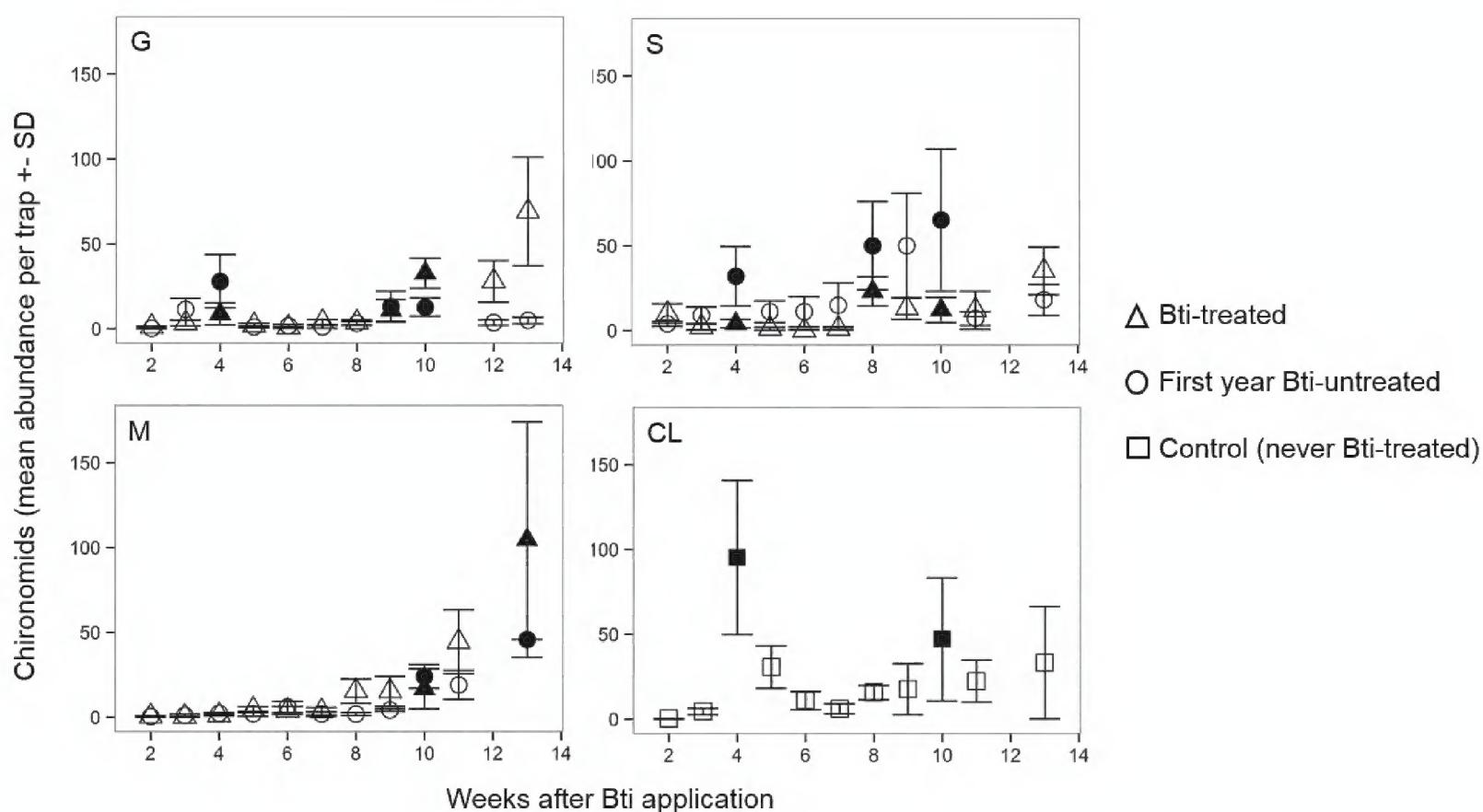


Figure 2. Chironomid mean abundances across all traps per site (M, G, S, and CL) for the whole sampling period. Different symbols refer to the varying Bti-treatments. Filled symbols indicate pooled emergence peak samples ($N = 18$) used for metabarcoding.

centration after clean-up was again measured using the Nanodrop and DNA concentrations of all samples were adjusted to $25 \text{ ng}/\mu\text{L}$.

A 322-bp fragment of the mitochondrial COI gene was amplified using the BF2 + BR1 primer set (Elbrecht et al. 2017). The used primer set was developed and evaluated with mock and *in silico* methods and does incorporate the needed degeneracy to amplify macroinvertebrates (including chironomids) reliably (Elbrecht and Leese 2017). The used fusion primers included Illumina adapter tails for sequencing (P5 or P7) and inline barcodes of different lengths for sample multiplexing (Elbrecht and Leese 2015). For each of the 18 samples two PCRs were conducted using the same primer pair but switching P5 and P7 Illumina adapters (Elbrecht and Leese 2015, Suppl. material 1). Sample 04CL was run with PCR replicates to test the PCR variability. PCR reactions consisted of $1\times$ PCR buffer (including 2.5 mM Mg^{2+}), 0.2 mM dNTPs , $0.5 \mu\text{M}$ of each primer, $0.025 \text{ U}/\mu\text{L}$ of HotMaster Taq (5Prime, Gaithersburg, MD, USA), 25 ng DNA , and HPLC H_2O to a total volume of $50 \mu\text{L}$. The PCR program included the following steps: 94°C for 3 minutes, 35 cycles of 94°C for 30 seconds, 50°C for 30 seconds, 65°C for 120 seconds and ended with 65°C for 5 minutes. PCR success was checked on a 1% agarose gel. Since some samples exhibited low DNA quantity (Qubit 2.0, Life Technologies, Carlsbad, CA, USA; measured concentration below $1 \text{ ng}/\mu\text{L}$), PCR for the respective samples was repeated with cycle number increased to 40 (Suppl. material 1). Amplicons were purified and size selected (retaining fragments of $>300 \text{ bp}$) with a left-sided size selection using magnetic beads (SpriSelect, Beckman Coulter, Bread, CA, USA, ratio: $0.76\times$). The DNA concentration was quantified using the Qubit and a high sensitivity (HS)

Assay Kit. Purified PCR products were pooled proportionately according to the number of specimens used in the extraction into a library to ensure all specimens are sequenced with the same sequencing depth. After pooling, the library was sent to an external laboratory (Macrogen, Seoul, Korea) for 300 bp paired-end sequencing on a MiSeq Illumina system (v3) run.

Bioinformatic analysis

Following the bioinformatic pipeline as previously described in Elbrecht and Leese (2017), the sequence data were processed as follows. In brief, after demultiplexing using a custom R script paired-end reads were merged to one sequence (Usearch version 8.8.1756). Primer sequences were removed via cutadapt (version 1.9.1). Singletons in each sample removed before clustering OTUs with the cluster_ots command at 97% identity Edgar 2013. All samples (including singletons) were matched against the OTUs (Usearch). To enhance data reliability, sequences matched to the respective OTU had to occur in both replicates and exceed the 0.003% threshold sequence abundance to be considered in downstream analysis (Elbrecht and Leese 2015). Finally, the obtained OTU sequences were matched against the BOLD database to retrieve taxon identification. All used metabarcoding pipeline and R scripts are available in Suppl. material 2.

Using metabarcoding data for chironomid diversity assessment

The retrieved chironomid species list was checked for biogeographic and ecologically plausibility, i.e., if the species names were listed for Germany and are repre-

sentative for temporary wetlands. If more than one species name per OTU was retrieved from BOLD with over 98% identity, we carefully examined the resulting hit table. For most hits, we then selected the biogeographically plausible species for our study region, based on the known biogeographical distribution and chironomid expert knowledge, for further ecological interpretations. If no clear decision could be made together with expert taxonomists, we followed the conservative approach to select the species name already represented in our data by another OTU. The species names retrieved in that way were categorized based on larval morphology in the context of standard water quality assessments into morphological determination “possible” (i.e., determination under 80× magnification without preparation), “difficult” (i.e., some characteristics need to be prepared and checked under greater magnification) and “impossible” (i.e., for species where the larva is not described, or do

not show morphological differences within one genus, or would demand highly elaborate preparation for species determination). Using this approach, we aimed to elucidate which proportion of the chironomid species pool is neglected in standard water quality assessments, where only the easily and quickly determinable chironomid larvae are considered. We then calculated the percentage of species retrieved via metabarcoding (here emergence data) in relation to species, which would have also been possible to determine morphologically in standard water quality assessments (usually larval data). Based on available saprobic indices of the chironomid species (Moog 1995, Moog 2002) retrieved via metabarcoding we also calculated the chironomid saprobic index (SI; Table 2) exemplarily for the four study sites across all WAA. When the same species name was retrieved from more than one OTU the respective abundances were summed up.

Table 1. Retrieved chironomid species names out of 54 obtained chironomid OTUs based on BOLD database searches. Given are OTU number(s), species names and the classification of the species determination based on larval morphology as routinely possible, difficult and impossible. Species names indicated with * are questionable.

| OTU | species names | morphological identification of larvae | | |
|--------------------------|--|--|-----------|------------|
| | | possible | difficult | impossible |
| OTU_15 | <i>Ablabesmyia monilis</i> (Linnaeus) | | × | |
| OTU_73 | <i>Acricotopus lucens</i> (Zetterstedt) | × | | |
| OTU_11 | <i>Chironomus annularius</i> (Meigen) | | | × |
| OTU_29 | <i>Chironomus curabilis*</i> (Bel et al.) | | | × |
| OTU_24 | <i>Chironomus melanescens</i> (Keyl) | | | × |
| OTU_5 | <i>Chironomus dorsalis</i> (Meigen) | | | × |
| OTU_13 + OTU_25 + OTU_12 | <i>Chironomus riparius</i> (Meigen) | × | | |
| OTU_42 | <i>Corynoneura scutellata</i> (Winnertz) | | | × |
| OTU_68 | <i>Corynoneura coronata</i> (Edwards) | | | × |
| OTU_35 | <i>Cricotopus sylvestris</i> (Fabricius) | | | × |
| OTU_40 | <i>Dicrotendipes lobiger</i> (Kieffer) | × | | |
| OTU_75 | <i>Limnophyes minimus</i> (Meigen) | | | × |
| OTU_18 + OTU_64 | <i>Limnophyes pentaplastus</i> (Kieffer) | | | × |
| OTU_34 | <i>Monopelopia tenuicalcar</i> (Kieffer) | × | | |
| OTU_47 | <i>Parachironomus parilis</i> (Walker) | | | × |
| OTU_4 | <i>Paralimnophyes longiseta</i> (Thienemann) | | | × |
| OTU_33 | <i>Paratanytarsus grimmii</i> (Schneider) | | | × |
| OTU_51 | <i>Paratanytarsus tenellulus</i> (Goetghebuer) | | | × |
| OTU_77 | <i>Paratendipes albimanus</i> (Meigen) | × | | |
| OTU_1 | <i>Polypedilum uncinatum</i> (Goetghebuer) | | | × |
| OTU_45 | <i>Procladius fuscus*</i> (Brundin) | | | × |
| OTU_19 + OTU_28 + OTU_97 | <i>Psectrocladius limbatus</i> (Holmgren) | | | × |
| OTU_32 | <i>Psectrotanytarsus varius</i> (Fabricius) | × | | |
| OTU_66 | <i>Rheocricotopus fuscipes</i> (Kieffer) | | × | |
| OTU_60 + OTU_264 | <i>Tanytarsus heusdensis</i> (Goetghebuer) | | × | |
| OTU_48 | <i>Tanytarsus pallidicornis</i> (Walker) | | × | |
| OTU_14 + OTU_137 | <i>Tanytarsus usmaensis</i> (Pagast) | | × | |
| OTU_7 | <i>Telmatopelopia nemorum</i> (Goetghebuer) | × | | |
| OTU_26 | <i>Xenopelopia nigricans</i> (Fittkau) | | | × |
| OTU_41 + OTU_79 | <i>Zavrelimyia barbatipes</i> (Kieffer) | | | × |
| N = 38 | N = 30 | N = 7 | N = 5 | N = 18 |

Table 2. Saprobi Index (SI) calculations per site ($N = 4$) across the whole sampling period based on 14 species retrieved from our data set for which the SI is available. Given are the species saprobi values (s), weights (w) as well as the species sequence frequencies (h) summed over all traps and sampling time points.

| Species names | s | w | h [G] | h [S] | h [M] | h [CL] |
|--|-----|---|--------|---------|--------|--------|
| <i>Ablabesmyia monilis</i> (Linnaeus) | 2.3 | 2 | 13 | 100,034 | 269 | 6 |
| <i>Chironomus riparius</i> (Meigen) | 3.5 | 3 | 21,341 | 63,995 | 47,395 | 75 |
| <i>Corynoneura scutellata</i> (Winnertz) | 1.7 | 2 | 80 | 17 | 2,622 | 237 |
| <i>Cricotopus sylvestris</i> (Fabricius) | 2.6 | 2 | 6 | 13,553 | 7 | 11 |
| <i>Limnophyes pentaplastus</i> (Kieffer) | 1.3 | 2 | 17 | 800 | 3 | 54,508 |
| <i>Monopelopia tenuicalcar</i> (Kieffer) | 0.8 | 4 | 8 | 6,828 | 0 | 0 |
| <i>Paratendipes albimanus</i> (Meigen) | 2.3 | 2 | 0 | 136 | 0 | 0 |
| <i>Psectrocladius limbatellus</i> (Holmgren) | 1.8 | 3 | 2,588 | 65,009 | 18 | 1,331 |
| <i>Psectrotanypus varius</i> (Fabricius) | 2.8 | 1 | 2,767 | 2,156 | 4,273 | 0 |
| <i>Rheocricotopus fuscipes</i> (Kieffer) | 2.2 | 3 | 0 | 0 | 0 | 120 |
| <i>Tanytarsus heusdensis</i> (Goetghebuer) | 1.4 | 1 | 4 | 857 | 0 | 0 |
| <i>Tanytarsus pallidicornis</i> (Walker) | 1.8 | 1 | 2 | 1,426 | 0 | 0 |
| <i>Xenopelopia nigricans</i> (Fittkau) | 0.8 | 2 | 403 | 114 | 16,913 | 0 |
| <i>Zavrelimyia barbatipes</i> (Kieffer) | 1.0 | 3 | 1 | 157 | 3,266 | 0 |
| SI | | | 3.3 | 2.5 | 2.8 | 1.3 |

Bti effects on chironomid community composition

To test whether the abundance data of emerged chironomids differed among Bti-treated and Bti-untreated sites (including the control site) for pooled samples of WAA 1 – WAA 4 (first emergence peak) and WAA 1 – WAA 13 (whole sampling period) a generalized linear mixed effect model (GLMM) (packages “nlme” v. 3.1-117, Pinheiro et al. 2016, and “MASS” v. 7.3-31, Venables and Ripley 2002) was implemented. As error structure the quasi Poisson family was chosen, where “study site” was implemented as random factor.

To test for differences in chironomid species richness between Bti-treated and Bti-untreated sites (including the control site), a Welch test was applied to the total number of OTUs and based on retrieved species names.

To test the hypothesis that the chironomid community composition differed between treatments, an adonis analysis (nonmetrical permutational MANOVA equivalent, Anderson 2001) was performed. For this, the Bray-Curtis distances of the Hellinger-transformed (see below) OTU read abundance assemblages per sites were calculated between pairs of sites, and these pairwise distances between sites were combined to a distance matrix of all sites, using the command ‘vegdist’ in the package “vegan” v. 2.4-1 (Oksanen et al. 2016). Within this distance matrix the nonmetrical permutational MANOVA equivalent was calculated using “Bti-treatment” as the distinguishing factor, with the command ‘adonis’ from the vegan package. We then assumed that time (WAA) would have a dominant effect on chironomid communities, but that Bti-treatment would alter the community as well. Therefore, also the interaction time * Bti-treatment was tested. Due to low number of replicates, the samples from the sites M and CL of WAA 8 and 9 were combined to the same time

period. For the sites S and G only WAA 8 and WAA 9, respectively, were available.

All analyses were conducted in R (<https://www.R-project.org>). For all multivariate analyses, the Hellinger transformation was chosen to give less weight to the few high abundant OTUs, since the abundance data were highly left-skewed with few taxa reaching abundances several orders of magnitude higher than those of the less abundant species (Legendre and Legendre 2012).

Results

Emergence data

In total, 11,589 emerged insects were collected, comprising of 17 taxa groups (Chironomidae: 78%; Culicidae: 14%; Trichoptera: 4%; Chaoboridae: 2%; Brachycera: 1%, other: 1%). On the Bti-treated sites 27 mosquito individuals were collected in the emergence traps, while on the Bti-untreated sites 1,006 mosquitoes emerged. Based on morphological identification 9,033 adult chironomids were collected. The number of chironomid specimens per emergence trap across all sites varied from 1 (Bti-treated site) to 1,239 (first year Bti-untreated site).

Emergence of chironomids fluctuated over time with varying emergence peaks at the Bti-untreated sites (Fig. 2). In particular, we detected one spring emergence peak (WAA 4) and two summer emergence peaks (WAA 9 + 10) at site G. At site S one spring peak (WAA 4) and two summer peaks (WAA 8 + 10) were detected. At site M, two summer peaks (WAA 10 + 13) were identified. A spring peak (WAA 4) and a summer peak (WAA 10) were identified at the never Bti-treated site CL. Specifically pooled emergence peak samples ($N = 18$, Figure

2) were selected for metabarcoding to investigate, if the abundance difference between Bti treated and untreated site pairs can be attributed to a shift in the chironomid community composition. The amount of individuals per pooled emergence peak sample varied from 22 to 541 (Suppl. material 1).

Bioinformatic analyses

In total 20,805,626 raw reads were generated by the MiSeq run with good read quality ($Q30 \geq 76.7\%$ of reads). Raw data are available at NCBI SRA archive (accession number SRR4244505; <https://www.ncbi.nlm.nih.gov/sra/?term=SRR4244505>). After demultiplexing, merging and trimming of PCR primers 8,869,048 sequences were used for further analysis. The number of sequences in each sample was significantly correlated ($p < 0.001$, adj. $R^2=0.942$) with the abundance of specimens per sample (Suppl. material 3).

OTU clustering analysis resulted in 442 OTUs. After application of the previously defined quality standards (0.003% minimum abundance) 89 OTUs were retained and used for subsequent analyses. The BOLD database searches identified 54 of the 89 OTUs (60.7%) as chironomids (Suppl. material 4). Of those, 38 OTUs (68.5%) could be assigned a species identification with 98 – 100% similarity, leaving 17 OTUs (31.5%) without species level identification (Suppl. material 4).

Using metabarcoding data for chironomid diversity assessment

In total 30 chironomid species were detected in the metabarcoding data set, with six species being assigned to 2–3 OTUs respectively (Table 1). For 11 OTUs we retrieved more than one species name with a sequence similarity of 97.82–100% (Suppl. material 4). For those OTUs, we were able to select the only biogeographically plausible species for our study region, based on biogeography and chironomid expert knowledge, for further ecological interpretations (underlined species names in Suppl. material 4). Only for OTU_12 two species names were biogeographically plausible, namely *Chironimus luridus* and *C. riparius*. Here, we selected the latter species as

C. riparius was also characterized by other OTUs in our data, whereas *C. luridus* was not represented otherwise.

Of the 30 retrieved species, seven species can be routinely determined (cost and time efficient) based on larval morphology in ecological water assessments, whereas the remaining species are difficult ($N = 5$) or impossible ($N = 18$) to determine based on larval morphology (Table 1). This resulted in a 73% increase of retrieved chironomid species names based on metabarcoding (using emergence data) in relation to morphological larvae determination.

The chironomid SI was calculated based on the available saprobic value for 14 chironomid species (45.2%) of our data set Table 2. The saprobic value per detected species ranged from 0.8 (*Monopelopia tenuicalcar*; *Xenopelopia nigricans*) up to 3.5 (*C. riparius*). For our study sites the chironomid SI ranged from 1.3 at the control site to 3.3 at the Bti treated sites (Table 2).

Bti effects on chironomid community composition

The abundance of emergent chironomids until WAA 4 at the Bti-treated sites was reduced by 64.99% compared to the abundance in the Bti-untreated sites (GLMM $t = 11.29$, $p = 0.008$, $df = 2$). After WAA 13, slightly more chironomids hatched at the Bti-treated vs. Bti-untreated sites (2,132 vs 1,800 individuals, respectively). However, this difference was not statistically significant (GLMM $t = -0.239884$, $df = 2$, $p = 0.833$).

Neither the number of OTU per sample (Welch two sample t-test, $t = 1.33$, $p = 0.20$) nor the number of species assigned from the OTU based on the data base (Welch two sample t-test, $t = 1.45$, $p = 0.17$) were significantly different between Bti-treated and Bti-untreated site pairs.

The adonis model of crossed Bti-treatment * site effects (with the variation from time implemented as groups (strata) within which permutations are constrained) explained 51% of the variation of the multivariate chironomid community composition, 34% of which were due to the differences within sites (Table 3). The effect from Bti treatment suggests a statistically significant, but only minor component explaining 12% of the variation ($p = 0.02$; Table 3).

Table 3. Results from the adonis analysis on the effect of treatment over time. The variation due to differences between sampling events was taken into account by the "strata = time" argument in the model. Df = degrees of freedom; F model = F statistic of the respective sub model.

| | Df | Sums of Squares | F Model | R ² | Pr(>F) | |
|-----------------------------|----|-----------------|---------|----------------|--------|-----|
| Site (time) | 3 | 0.82 | 1.89 | 0.34 | 0.003 | *** |
| Bti Treatment | 1 | 0.30 | 2.05 | 0.12 | 0.02 | * |
| Site (time) : Bti-Treatment | 2 | 0.13 | 0.90 | 0.05 | 0.45 | |
| Residuals | 8 | 1.15 | | 0.48 | | |
| Total | 13 | 2.40 | | 1.00 | | |

Discussion

Using metabarcoding data for chironomid diversity assessment

With our metabarcoding approach we detected 54 chironomid OTUs across all study sites, of which almost 70% could be identified to species level using the BOLD database. Even though we did not have a specific reference database for our study system (e.g. Carew et al. 2013), we have mainly extracted biogeographically and ecologically meaningful species names as many of these species are frequently found in periodically desiccative ponds as euryoecious ubiquists (e.g., *Ablabesmyia monilis*, *Acricotopus lucens*, *Chironomus riparius*, *Corynoneura scutellata*, *Cricotopus sylvestris*, *Limnophyes pentaplastus*, *Paratendipes albimanus*, *Polydendrum uncinatum*, *Psectrotanytarsus varius* and *Tanytarsus pallidicornis*). Despite selecting only chironomid specimens for metabarcoding, also non-chironomid OTUs ($N = 35$; 39.3%) were detected in low abundances (Suppl. material 4). These records included other Dipterans ($N = 14$; 15.7%), Trichopterans ($N = 2$; 2.2%), Lepidopterans ($N = 2$; 2.2%), Arachnids (Pionidae: $N = 6$; 6.7%), Fungi (Sporidiobolale, $N = 2$, Eurotiales, $N = 1$, Tremellales, $N = 1$ and Microstromatales, $N = 1$; 5.6% in total) and Bacteria (Rickettsia: $N = 1$; 1.1%). In addition, OTUs without hit in the BOLD-database ($N = 6$; 6.7%) were detected. However, the presence of other taxa in the data is most likely related to the universality of the primer pair used, which could have amplified traces of other taxa previously stored in the same collection tubes, thus causing the large number of low level OTUs detected. Previous experiences show that low abundance OTUs are often derived from PCR or sequencing errors as well as chimeras and non-target DNA of small organisms (Elbrecht and Leese 2015, 2017, Elbrecht et al. 2017). As DNA was extracted from tissue bulk samples, eDNA and small non-target specimens are not of interest for this study. Thus, the data was not further analyzed. It should also be noted that the lower the OTU abundance is the more there are stochastic effects between samples, making it difficult to analyze OTUs with very low abundance.

For 11 OTUs we retrieved more than one species name with a sequence similarity of 97.82–100 % (Suppl. material 4). This could indicate 1) limited taxonomic resolution of the short fragment amplified by the used primer set, producing only 322 bp COI fragments opposed to the 658 bp COI fragments using the classical Folmer primers (Folmer 1994); 2) different taxonomic keys used thus having different synonyms included; or 3) potential taxonomic misidentification which are also discussed in (Elbrecht et al. 2017) who recommend better data curation in taxonomic databases. In contrast, six chironomid species names were retrieved from two or three different OTUs, respectively, and two OTUs (45, 29) were assigned to questionable species names due to their biogeography (*Procladius cf. fuscus* and *C. curabilis*, Table 1). This could suggest

cryptic intraspecific diversity, as we used a species divergence rate of 3% which might be too low for some species (Ekrem et al. 2007, Carew et al. 2013). As little is known about the genetic lineage of Chironomidae, cryptic species or variable phenotypes could be possible and hamper the correct identification by taxonomist (Anderson et al. 2013, Carew et al. 2007, Stur and Ekrem 2011).

The information benefit of metabarcoding by obtaining species names strongly depends on the quality of the database. For 17 chironomid OTUs (31.5%) no species identification could be obtained (Suppl. material 4). BOLD holds 270,292 published records of Chironomidae forming 5,540 BINs (clusters) with specimens from 49 countries. Of these records (accessed on 14.07.2017), 100,231 have species names, and represent 1,233 species for an estimated species diversity of 15,000 worldwide (Armitage et al. 1995). For Germany, with an estimated species richness of approx. 700 different Chironomidae (Samietz 1996), BOLD has 3,706 published records forming 217 BINs (clusters). Of these records (accessed on 14.07.2017), 3,683 have species names, representing only 208 species (around 30%). Metabarcoding can only be as good as the database on which it relies for OTU matching to species identifications. We thus encourage experienced chironomid taxonomists to increase the number chironomid species in the BOLD database to even improve the effectiveness of metabarcoding for chironomid diversity assessments.

By applying metabarcoding we obtained 70% more chironomid species identifications than would have been possible based on traditional taxonomic determination of larval samples (Table 1), thus proving the usefulness of metabarcoding for chironomid diversity assessment. Some of our retrieved species are indicators for high water quality and were previously detected in spring biotopes, such as *Acricotopus lucens*, *Chironomus liridus*, *Dicrotendipes lobiger*, *Limnophyes minimus*, *Limnophyes pentaplastus*, *Psectrocladius limbatellus*, *Psectrotanytarsus varius* and *Tanytarsus usmaensis* (Reiff et al. 2015), suggesting a general good water quality of our study sites. Only seven of those species can be determined based on larval morphology (Table 1). Even though various determination keys for larval and adult chironomids exist, not all taxa can be determined to species level even by experts (Kranzfelder et al. 2016). Especially larvae and female midges are almost impossible to determine morphologically, since often male genitalia are necessary to distinguish species. Some chironomid species have a parthenogenetic life cycle (e.g. *Paratanytarsus grimmii*, Langton 1988), so only females occur especially in temporary wetlands (Dettinger-Klemm 2003). Without determination of all occurring chironomids, including females, around 27% of the species diversity could be lost (Ekrem et al. 2010). Even if morphology enables the determination down to genus level, an ecological interpretation is difficult since chironomid species of the same genus might have very dissimilar ecological preferences Milošević et al. (2013).

The saprobic index per site based on 14 chironomid species and their sequence abundances ranged from 1.3 (control site) to 3.3 in one of the Bti treated sites (Table 2). Due to the demanding morphological chironomid species determination, it is common practise to exclude chironomids from bioassessment programs (Milošević et al. 2013). However, in standard water quality assessments in Germany sometimes only all red chironomid larvae are counted, summarized as *Chironomus spec.* and included in the saprobic index with a value around 3.5. Considering the high chironomid diversity and the range of saprobic values for chironomids between 0.8 (very good water quality) and 3.5 (bad water quality), the standard water quality assessment would have resulted in a severe underestimation of the studied water bodies due to the presence of *C. riparius*. In addition to the difficult morphological determination, small chironomid larvae (< 1 mm) from freshly hatched species can be easily overlooked by larvae picking. Thus, a metabarcoding approach based on water and homogenized sediment samples could be highly useful for future application in water quality assessments by increasing the chironomid diversity in a sample without specialised taxonomic expertise needed (Bista et al. 2017).

The advantages of metabarcoding over traditional monitoring for water quality assessments is gaining increasing attention. Since Haase et al. 2010 postulated the overlooking of many taxa in traditional stream monitoring programs, many studies proved that metabarcoding can provide higher numbers and more accurate taxonomic identifications than morphology-based methods for many freshwater macroinvertebrates (Hajibabaei et al. 2011, Carew et al. 2013, Elbrecht and Leese 2015, Elbrecht et al. 2017). Moreover, barcoding has increased for rapid biodiversity assessment and biomonitoring for many terrestrial taxa (Yu et al. 2012, Taberlet et al. 2012, Brehm et al. 2013). Ji et al. (2013) compared metabarcoded samples of arthropods and birds with standard biodiversity data sets, and found that the genetic data sets were taxonomically more comprehensive, quicker to produce and less reliant on taxonomic expertise. Cristescu (2014) raised the urge for a coordinated progression of species barcoding that integrates taxonomic expertise and genetic data. For the family Chironomidae an extended and reliably curated barcode database (analogous to the Trichoptera Barcode of Life Database, Zhou et al. 2016) would be highly useful for integrating chironomids in standard freshwater biomonitoring which enhance water quality assessments and might lead to better management of aquatic ecosystems.

Bti effects on chironomid community composition

In our study sites we could show that a considerable number of chironomids live in these wetlands subjected to mosquito control. The uptake and the mode of action of Bti is similar for mosquitos and chironomids (Ali et al. 1981). Regarding potential effects of mosquito control

actions using the biocide Bti we expected an overall reduction in chironomid abundance in the Bti-treated sites as well as a reduction in species richness and resulting community composition changes.

The chironomid abundance until WAA 4 was significantly reduced by almost 65% in the Bti-treated sites compared to the Bti-untreated sites, including the never Bti-treated control site. At the control site this spring peak was especially pronounced, indicating that WAA 4 (here: begin of May) is a key time period for the overall chironomid emergence in this area. The observed abundance reduction can be explained by the recent Bti treatment, which killed not only the mosquito larvae but also the chironomid larvae, predominantly affecting freshly hatched larvae. Especially first instar larvae of *C. riparius* were shown to be highly affected by Bti in laboratory experiments while older larvae were less sensitive (Kästel et al. 2017). Until WAA 13 there was a non-significant trend towards more chironomids in the Bti-treated sites, which could be due to a reduced mosquito competition (Lundström et al. 2010) and subsequently chironomids with a second reproductive cycle in the same year had better conditions (more food resources available) to reproduce. Moreover, species have different egg laying and hatching times, and Bti does not affect eggs but only hatched individuals (Boisvert and Boisvert 2000). The species richness, however, was not significantly different between Bti-treated and untreated sites, neither on OTU-level, accounting for potential cryptic species diversity, nor on species level. The Bti-treatment thus seems to have a mainly quantitative effect on the abundance of the species present in the communities, which is stronger shortly after application during the main chironomid emergence peak in spring.

The adonis analysis corroborated the assumption that site and time (seasonality) had a dominant effect on chironomid communities (Table 3). The predominant effect of site can be explained by the study sites different vegetation (grassland, alder carr, oak carr and pine forest) which influences species composition by varying substrate availability, water chemistry, and the availability of nutritional resources (Van Den Brink and Van Der Velde 1991). Thus, it is not feasible to directly compare the chironomid species composition on the never Bti-treated control site (CL) and the Bti-treated sites (S, G and M) among each other since the vegetational surroundings are quite different and so is the species composition (see Table 2). Note, however, that it is hardly possible to find a “true” control site in the Upper Rhine Valley, i.e., a wetland which has never been treated with Bti next to wetlands subject to mosquito control. In this study we have therefore compared four different study sites, three of which have been subject to mosquito control with Bti.

Even though site and time influenced species composition the most, the first year of Bti intermittence significantly altered the chironomid community as well. This Bti effect was rather low (12%, Table 3). However, considering the more than 20 years of continued Bti application in

the study area each spring (in some years even several applications per season) and the proven toxic effect of Bti on chironomid first instar larvae (Kästel et al. 2017), we can assume a more or less depleted community in terms of chironomid diversity. As dispersal for adult chironomids from the next Bti-untreated areas might take too long given the reduced flight capacities (Armitage et al. 1995), a recolonization of univoltine species would probably need longer than one season intermitting Bti treatment. Therefore, resilience in terms of significantly increased species richness may even only be expected after several seasons intermitting of the Bti treatment. This highlights the importance for follow-up studies at the sites.

Conclusions

We showed the effectiveness of metabarcoding for chironomid diversity assessments, which led to a 70% increase in species determination compared to determination based on larval morphology. Thus, metabarcoding improves data quality by generating taxonomic resolution. Regarding the question of non-target effects of Bti on the chironomid community, our study found only minor significant effects even though Bti reduced the chironomid emergence by 65%. This could be due to a time lag of chironomid recolonization, since the study year was the first year of Bti intermittence after around 20 years of Bti application in the study area. A follow-up study after a few years of Bti intermittence could result in a more obvious recovery of the chironomid community composition in the Bti-untreated temporary wetlands.

Acknowledgements

This work has been financed by the Ministerium für Wissenschaft, Weiterbildung und Kultur Rheinland-Pfalz, Germany, in the frame of the programme “Research initiative”, project AufLand. We are grateful to the representatives of the town Neustadt an der Weinstraße and to the Stiftung Natur und Umwelt Rheinland-Pfalz for supporting this project ideally and financially. FL and VE are supported by the EU COST (European Cooperation in Science and Technology) Action DNAqua-Net (CA15219). We thank Martin Spies, Zoologische Staats-sammlung München, for advices with questionable species and Christoph Leeb for generating the map of the study sites. Furthermore, we would like to thank Torbjørn Ekrem, Iliana Bista and Kristy Deiner for valuable comments substantially improving this manuscript.

Authors' contributions: Sampling: AK, SA; Laboratory work: AK, VE; Bioinformatic data analyses: VE; Biological data analyses: KT, AK, JM, SM; Multivariate statistics: SIS; Study design and supervision: KT; CB; FL; Manuscript writing: KT; AK; equal contribution.

All authors edited and commented on the manuscript draft.

References

- Ali A, Baggs RD, Stewart JP (1981) Susceptibility of some Florida chironomids and mosquitoes to various formulations of *Bacillus thuringiensis* var. *israelensis*. Journal of Economic Entomology 74: 672–677. <https://doi.org/10.1093/jee/74.6.672>
- Aljanabi S, Martinez I (1997) Universal and rapid salt-extraction of high quality genomic DNA for PCR-based techniques. Nucleic Acids Research 25 (22): 4692–4693. <https://doi.org/10.1093/nar/25.22.4692>
- Anderson AM, Stur E, Ekrem T (2013) Molecular and morphological methods reveal cryptic diversity and three new species of Nearctic Micropsectra (Diptera: Chironomidae). Freshwater Science 32 (3): 892–921. <https://doi.org/10.1899/12-026.1>
- Anderson M (2001) Permutation tests for univariate or multivariate analysis of variance and regression. Canadian Journal of Fisheries and Aquatic Science. <https://doi.org/10.1139/cjfas-58-3-626>
- Armitage PD, Cranston PS, Pinder LC (1995) The Chironomidae: Biology and Ecology of Non-Biting Midges. Chapman and Hall, London. <https://doi.org/10.1007/978-94-011-0715-0>
- Batzer D, Boix D (Eds) (2016) Invertebrates in Freshwater Wetlands: An International Perspective on Their Ecology. Springer, Cham. <https://doi.org/10.1007/978-3-319-24978-0>
- Becker N (1998) The use of *Bacillus thuringiensis* subsp. *israelensis* (Bti) against mosquitoes, with special emphasis on the ecological impact. Israel Journal of Entomology 63–69.
- Biggs J, Williams P, Whitfield M, Nicolet P, Weatherby A (2005) 15 years of pond assessment in Britain: results and lessons learned from the work of Pond Conservation. Aquatic Conservation: Marine and Freshwater Ecosystems 15: 693–714. <https://doi.org/10.1002/aqc.745>
- Bista I, Carvalho G, Walsh K, Seymour M, Hajibabaei M, Lallias D, Christmas M, Creer S (2017) Annual time-series analysis of aqueous eDNA reveals ecologically relevant dynamics of lake ecosystem biodiversity. Nature Communications 8: 14087. <https://doi.org/10.1038/ncomms14087>
- Boisvert M, Boisvert J (2000) Effects of *Bacillus thuringensis* var. *israelensis* on target and nontarget organisms: a review of laboratory and field experiments. Biocontrol Science and Technology 10 (5): 517–561. <https://doi.org/10.1080/095831500750016361>
- Brehm G, Strutzenberger P, Fielder K (2013) Phylogenetic diversity of geometrid moths decreases with elevation in the tropical Andes. Ecography 36: 1247–1253. <https://doi.org/10.1111/j.1600-0587.2013.00030.x>
- Carew M, Pettigrove V, Cox R, Hoffmann A (2007) The response of Chironomidae to sediment pollution and other environmental characteristics in urban wetlands. Freshwater Biology 52 (12): 2444–2462. <https://doi.org/10.1111/j.1365-2427.2007.01840.x>
- Carew M, Pettigrove V, Metzeling L, Hoffmann A (2013) Environmental monitoring using next generation sequencing: rapid identification of macroinvertebrate bioindicator species. Frontiers in Zoology 10: 45. <https://doi.org/10.1186/1742-9994-10-45>
- Cristescu M (2014) From barcoding single individuals to metabarcoding biological communities: Towards an integrative approach to the study of global biodiversity. Trends in Ecology and Evolution 29 (10): 566–571. <https://doi.org/10.1016/j.tree.2014.08.001>
- Dettinger-Klemm P-A (2003) Chironomids (Diptera, Nematocera) of temporary pools - an ecological case study. PhD Thesis, University Heidelberg, Heidelberg.

Edgar RC (2013) UPARSE: Highly accurate OTU sequences from microbial amplicon reads. *Nature Methods* 10: 996–998. <https://doi.org/10.1038/nmeth.2604>

Ekrem T, Stur E, Hebert P (2010) Females do count: documenting Chironomidae (Diptera) species diversity using DNA barcoding. *Organisms Diversity and Evolution* 10: 397–408. <https://doi.org/10.1007/s13127-010-0034-y>

Ekrem T, Willassen E, Stur E (2007) A comprehensive DNA sequence library is essential for identification with DNA barcodes. *Molecular Phylogenetics and Evolution* 43: 530–542. <https://doi.org/10.1016/j.ympev.2006.11.021>

Elbrecht V, Leese F (2015) Can DNA-based ecosystem assessments quantify species abundance? Testing primer bias and biomass - sequence relationships with an innovative metabarcoding protocol. *PLoS One* 10 (7): e0130324. <https://doi.org/10.1371/journal.pone.0130324>

Elbrecht V, Leese F (2017) Validation and development of COI metabarcoding primers for freshwater macroinvertebrate bioassessment. *Frontiers in Environmental Sciences* 5: 1–16. <https://doi.org/10.3389/fenvs.2017.00011>

Elbrecht V, Vamos EE, Meissner K, Aroviita J, Leese F (2017) Assessing strengths and weaknesses of DNA metabarcoding-based macroinvertebrate identification for routine stream monitoring. *Methods in Ecology and Evolution* 8 (10): 1265–1275. <https://doi.org/10.1111/2041-210X.12789>

Ferrington L (2008) Global diversity of non-biting midges (Chironomidae; Insecta-Diptera) in freshwater. *Hydrobiologia* 595: 447–455. https://doi.org/10.1007/978-1-4020-8259-7_45

Folmer O (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3: 294–299.

Gibson J, Shokralla S, Curry C, Baird D, Monk W, King I, Hajibabaei M (2015) Large-scale biomonitoring of remote and threatened ecosystems via high-throughput sequencing. *PLoS One* 10 (10): e0138432. <https://doi.org/10.1371/journal.pone.0138432>

Haase P, Pauls S, Schindelhütte K, Sundermann A (2010) First audit of macroinvertebrate samples from an EU Water Framework Directive monitoring program: human error greatly lowers precision of assessment results. *Journal of the North American Benthological Society* 29 (4): 1279–1291. <https://doi.org/10.1899/09-183.1>

Hajibabaei M, Shokralla S, Zhou X, Singer GG, Baird D (2011) Environmental Barcoding: A Next-Generation Sequencing Approach for Biomonitoring Applications Using River Benthos. *PLoS One* 6 (4): e17497. <https://doi.org/10.1371/journal.pone.0017497>

Hebert PN, Ratnasingham S, de Waard J (2003) Barcoding Animal Life: Cytochrome c Oxidase Subunit 1 Divergences among Closely Related Species. *Proceedings of the Royal Society of London B: Biological Sciences* 270: 96–99. <https://doi.org/10.1098/rsbl.2003.0025>

Hershey A, Shannon L, Axler R, Ernst C, Mickelson P (1995) Effects of methoprene and *Bti* (*Bacillus thuringiensis* var.*israelensis*) on non-target insects. *Hydrobiologia* 308: 219–227. <https://doi.org/10.1007/BF00006873>

Jakob C, Poulin B (2016) Indirect effects of mosquito control using *Bti* on dragonflies and damselflies (Odonata) in the Camargue. *Insect Conservation and Diversity* 9: 161–169. <https://doi.org/10.1111/icad.12155>

Ji Y, Ashton L, Pedley S, Edwards D, Tang Y, Nakamura A, Kitching R, Dolman P, Woodcock P, Edwards F, Larsen T, Hsu W, Benedict S, Hamer K, Wilcove D, Bruce C, Wang X, Levi T, Lott M, Emerson B, Yu D (2013) Reliable, verifiable and efficient monitoring of biodiversity via metabarcoding. *Ecology Letters* 16: 1245–1257. <https://doi.org/10.1111/ele.12162>

Kästel A, Allgeier S, Brühl C (2017) Decreasing *Bacillus thuringiensis israelensis* sensitivity of *Chironomus riparius* larvae with age indicates potential environmental risk for mosquito control. *Scientific Reports* 7: 13565. <https://doi.org/10.1038/s41598-017-14019-2>

Kranzfelder P, Ekrem T, Stur E (2016) Trace DNA from insect skins: a comparison of five extraction protocols and direct PCR on chironomid pupal exuviae. *Molecular Ecology Resources* 16 (1): 353–363. <https://doi.org/10.1111/1755-0998.12446>

Kwong S, Srivathsan A, Meier R (2012) An update on DNA barcoding: low species coverage and numerous unidentified sequence. *Cladistics* 28: 639–644. <https://doi.org/10.1111/j.1096-0031.2012.00408.x>

Lagadic L, Schäfer RB, Roucaute M, Szöcs E, Chouin S, de Maupeou J, Duchet C, Franquet E, Hunsec B, Bertrand C, Fayolle S, Francés B, Rozier Y, Foussadier R, Santoni J, Lagneau C (2016) No association between the use of *Bti* for mosquito control and the dynamics of non-target aquatic invertebrates in French coastal and continental wetlands. *Science of The Total Environment* 553: 486–494. <https://doi.org/10.1016/j.scitotenv.2016.02.096>

Langton PH (1988) The parthenogenetic midge of water supply systems, *Paratanytarsus grimmii* (Schneider) (Diptera: Chironomidae). *Bulletin of Entomological Research* 78: 317–328. <https://doi.org/10.1017/S0007485300013080>

Legendre P, Legendre L (2012) Numerical ecology. Elsevier, Amsterdam.

Liber K, Schmude K, Rau D (1998) Toxicity of *Bacillus thuringiensis israelensis* var. *israelensis* to chironomids in pond mesocosms. *Ecotoxicology* 7: 343–354. <https://doi.org/10.1023/A:1008867815244>

Lindegaard C, Brodersen K (1995) Distribution of Chironomidae (Diptera) in the river continuum. In: Cranston P (Ed.) Chironomids: from genes to ecosystems. CSIRO, Melbourne, 257–271 pp.

Lukács B, Sramkó G, Molnár A (2013) Plant diversity and conservation value of continental temporary pools. *Biological Conservation* 158: 393–400. <https://doi.org/10.1016/j.biocon.2012.08.024>

Lundström J, Schäfer M, Petersson E, Persson-Vinnersten T, Landin J, Brodin Y (2010) Production of wetland Chironomidae (Diptera) and the effects of using *Bacillus thuringiensis israelensis* for mosquito control. *Bulletin of Entomological Research* 100 (01): 117–125. <https://doi.org/10.1017/S0007485309990137>

Milošević D, Simić V, Stojković M, Čerba D, Mančev D, Petrović A, Paunović M (2013) Spatio-temporal pattern of the Chironomidae community: Toward the use of non-biting midges in bioassessment programs. *Aquatic Ecology* 47 (1): 37–55. <https://doi.org/10.1007/s10452-012-9423-y>

Moog O (1995) Fauna Aquatica Austriaca. Wasserwirtschaftskataster, Bundesministerium f. Land- und Forstwirtschaft, Wien.

Moog O (2002) Fauna Aquatica Austriaca, Katalog zur autökologischen Einstufung aquatischer Organismen Österreichs. Wasserwirtschaftskataster, Bundesministerium für Land- und Forstwirtschaft, Umwelt und Wasserwirtschaft, Wien.

Niemi G, Hershey A, Shannon L, Hanowski J, Lima A, Axler R, Regal R (1999) Ecological effects of mosquito control on zooplankton, insects, and birds. *Environmental Toxicology and Chemistry* 18 (3): 549–559. <https://doi.org/10.1002/etc.5620180325>

Oksanen FGB, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara RB, Simpson GL, Solymos P, Stevens MHH, Szoeics E,

Wagner H (2016) vegan: Community Ecology Package. R package. 2.4-1. URL: <https://CRAN.R-project.org/package=vegan>

Pfitzner W, Beck M, Weitzel T, Becker N (2015) The role of mosquitoes in the diet of adult dragon and damselflies (Odonata). Journal of American Mosquito Control Association 31 (2): 187–189. <https://doi.org/10.2987/14-6476R>

Ping L, Wen-Ming Z, Shui-Yun Y, Jin-Song Z, Li-Jun L (2005) Impact of environmental factors on the toxicity of *Bacillus thuringiensis* var. *israelensis* ips82 to *Chironomus kiiensis*. Journal of American Mosquito Control Association 21: 59–63. [https://doi.org/10.2987/8756-971X\(2005\)21\[59:IOEFOT\]2.0.CO;2](https://doi.org/10.2987/8756-971X(2005)21[59:IOEFOT]2.0.CO;2)

Pinheiro J, Bates D, DebRoy S, Sarkar D, Team RC (2016) nlme: linear and nonlinear mixed effects models. R package. 3.1-128. <http://CRAN.R-project.org/package=nlme>

Poulin B, Levebre G, Paz L (2010) Red flag for green spray: adverse trophic effects of *Bti* on breeding birds. Journal of Applied Ecology 47 (4): 884–889. <https://doi.org/10.1111/j.1365-2664.2010.01821.x>

Puntí T, Rieradevall M, Prat N (2009) Environmental factors, spatial variation, and specific requirements of Chironomidae in Mediterranean reference streams. Journal of the North American Benthological Society 28: 247–265. <https://doi.org/10.1899/07-172.1>

Rathnasingham S, Hebert P (2007) BOLD: The Barcode of Life Data System (www.barcodinglife.org). Molecular Ecology Notes 7: 355–364. <https://doi.org/10.1111/j.1471-8286.2006.01678.x>

Raunio J, Heino J, Paasivirta L (2011) Non-biting midges in biodiversity conservation and environmental assessment: findings from boreal freshwater ecosystems. Ecological Indicators 11: 1057–1064. <https://doi.org/10.1016/j.ecolind.2010.12.002>

Reiff N, Michiels S, Reusch H, Wagner R, Gerecke R (2015) Die Dipteren-Fauna der Kleingewässer im Berggrutschgebiet Hirschkopf bei Mössingen (Landkreis Tübingen, Baden-Württemberg), unter besonderer Berücksichtigung der Chironomidae. Lauterbornia 80: 1057–1064.

Rodcharoen J, Mulla M, Chaney J (1991) Microbial larvicides for the control of nuisance aquatic midges (Diptera: Chironomidae) inhabiting mesocosms and man-made lakes in California. Journal of the American Mosquito Control Association 7 (1): 56–62.

Rossaro B, Lencioni V, Boggero A, Marziali L (2006) Chironomids from Southern Alpine running waters: ecology, biogeography. Hydrobiologia 562: 231–246. <https://doi.org/10.1007/s10750-005-1813-x>

Samietz R (1996) Kommentiertes Verzeichnis der auf dem Gebiet der Bundesrepublik Deutschland nachgewiesenen Chironomiden-Arten (Insecta; Diptera). Abhandlungen und Berichte des Museums der Natur Gotha 19: 36–70.

Stav G, Blaustein L, Margalit Y (2005) Individual and interactive effects of a predator and con-trophic species on mosquito populations. Ecological Applications 15 (2): 587–598. <https://doi.org/10.1890/03-5191>

Sæther OA (1979) Chironomid communities as water quality indicators. Ecography 2 (2): 65–74. <https://doi.org/10.1111/j.1600-0587.1979.tb00683.x>

Stur E, Ekrem T (2011) Exploring unknown life stages of Arctic Tanytarsini (Diptera: Chironomidae) with DNA barcoding. Zootaxa 2743: 27–39.

Sweeney B, Battle J, Jackson J, Dapkey T (2011) Can DNA barcodes of stream macroinvertebrates improve descriptions of community structure and water quality? Journal of the North American Benthological Society 30: 195–216. <https://doi.org/10.1899/10-016.1>

Taberlet P, Coissac E, Pompanon F, Brochmann C, Willerslev E (2012) Towards next-generation biodiversity assessment using DNA metabarcoding. Molecular Ecology 21 (8): 2045–2050. <https://doi.org/10.1111/j.1365-294X.2012.05470.x>

Treverrow N (1985) Susceptibility of *Chironomus tepperi* (Diptera: Chironomidae) to *Bacillus thurengiensis* serovar *israelensis*. Journal of the Australian Entomological Society 303–304. <https://doi.org/10.1111/j.1440-6055.1985.tb00248.x>

Van Den Brink FWB, Van Der Velde B (1991) Macrozoobenthos of floodplain waters of the rivers rhine and meuse in the Netherlands: A structural and functional analysis in relation to hydrology. River Research and Applications 6 (4): 265–277. <https://doi.org/10.1002/rrr.3450060405>

Vaughan IP, Newberry C, Hall DJ, Liggett JS, Ormerod SJ (2008) Evaluating large-scale effects of *Bacillus thuringiensis* var. *israelensis* on non-biting midges (Chironomidae) in a eutrophic urban lake. Freshwater Biology 53 (10): 2117–2128. <https://doi.org/10.1111/j.1365-2427.2008.02043.x>

Venables WN, Ripley BD (2002) Modern Applied Statistics with R. 4. Springer, NY. <https://doi.org/10.1007/978-0-387-21706-2>

Williams D (2006) The biology of temporary waters. Oxford University Press, Oxford.

Yiallouros M, Storch V, Becker N (1999) Impact of *Bacillus thuringiensis* var. *israelensis* on larvae of *Chironomus thummi thummi* and *Psectrocladius psilopterus* (Diptera: Chironomidae). Journal of Invertebrate Pathology 74: 39–47. <https://doi.org/10.1006/jipa.1999.4852>

Yu D, Ji Y, Emerson B, Wang X, Ye C, Yang C, Ding Z (2012) Biodiversity soup: Metabarcoding of arthropods for rapid biodiversity assessment and biomonitoring. Methods in Ecology and Evolution 3 (4): 613–623. <https://doi.org/10.1111/j.2041-210X.2012.00198.x>

Zhou X, Frandsen P, Holzenthal R, Beet C, Bennett K, Blahnik R, Bonada N, Cartwright D, Chuluunbat S, Cocks G, et al. (2016) The Trichoptera barcode initiative: a strategy for generating a species-level Tree of Life. Philosophical Transactions B 371: 20160025. <https://doi.org/10.1098/rstb.2016.0025>

Supplementary material 1

Sample ID, specimen number, DNA and amplion concentrations, primer combination, number of PCR cycles and final column for library preparation

Authors: Kathrin Theissinger, Anna Kästel, Vasco Elbrecht, Jenny Makkonen, Susanne Michiels, Susanne I. Schmidt, Stefanie Allgeier, Florian Leese and Carsten Brühl

Data type: excel table

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mbmg.2.21060.suppl1>

Supplementary material 2**Bioinformatic pipeline**

Authors: Vasco Elbrecht

Data type: R scripts

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mbmg.2.21060.suppl2>

Link: <https://doi.org/10.3897/mbmg.2.21060.suppl3>

Supplementary material 4**OTU table**

Authors: Kathrin Theissinger, Anna Kästel, Vasco Elbrecht, Jenny Makkonen, Susanne Michiels, Susanne I. Schmidt, Stefanie Allgeier, Florian Leese and Carsten Brühl

Data type: excel spread sheet

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mbmg.2.21060.suppl4>

Supplementary material 3**Number of specimens per sample as a function of the number of sequences per sample**

Authors: Kathrin Theissinger, Anna Kästel, Vasco Elbrecht, Jenny Makkonen, Susanne Michiels, Susanne I. Schmidt, Stefanie Allgeier, Florian Leese and Carsten Brühl

Data type: Text and Figure

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.